



CASE REPORT

When Chromosome 15 Duplicates: A Clinical-Genetic Challenge

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Abstract

Background: Chromosome 15q11–q13 duplication syndrome (Dup15q) is a rare neurodevelopmental disorder caused by additional copies of a genomic region enriched in imprinted genes essential for normal neuronal function. It is commonly associated with autism spectrum disorder (ASD), epilepsy, hypotonia, global developmental delay, and intellectual disability. **Case presentation:** We report the case of a 31-year-old male with a longstanding history of refractory epilepsy, autism spectrum disorder, and intellectual disability. Genetic evaluation using array-comparative genomic hybridization (array-CGH) combined with single-nucleotide polymorphism (SNP) analysis identified a de novo interstitial heterozygous duplication of 10.03 Mb involving the 15q11.2–q13.3 region. The duplicated segment encompasses multiple genes with key roles in neurodevelopment, including UBE3A, GABRB3, GABRA5, SNRPN, and NDN. Parental testing by multiplex ligation-dependent probe amplification (MLPA) was negative, confirming a non-inherited origin. Establishing the molecular diagnosis allowed precise clinical classification, informed therapeutic decision-making, and enabled appropriate genetic counseling. **Conclusion:** This case illustrates the diagnostic value of array-CGH in patients with complex neurodevelopmental phenotypes and emphasizes the importance of early genetic assessment in individuals presenting with ASD and epilepsy. Identification of pathogenic copy number variants such as Dup15q has significant implications for prognosis, clinical management, and family counseling.

INTRODUCTION

15q11.2–q13.3 duplication syndrome (Dup15q) is a rare neurodevelopmental disorder with autosomal dominant inheritance caused by duplication of the chromosomal segment 15q11.2–q13.3. This region, located on the long arm of

chromosome 15, is highly susceptible to recombination errors during meiosis due to the presence of low-copy repeat sequences (LCRs). These structural anomalies arise primarily through non-allelic homologous recombination (NAHR) between specific breakpoint regions (BP1 to BP5) [1].

The duplication may occur in two main forms: as an interstitial duplication on the maternal chromosome 15 or as a supernumerary isodicentric chromosome [idic(15)], which involves two additional copies of the duplicated region. These variants confer variable clinical expressivity and incomplete penetrance; some carriers may be asymptomatic or present with a mild phenotype. Maternal-origin duplications have been associated with more severe clinical manifestations compared to paternal-origin ones [1].

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From a pathophysiological perspective, it has been proposed that the gene dosage imbalance caused by the duplication contributes to pathological overexpression of genes critical to brain development. This phenomenon may interfere with synaptic maturation, neuronal plasticity, and ion channel regulation—processes essential for proper central nervous system function. These neurobiological alterations underlie the hallmark signs and symptoms of the syndrome [2].

Clinically, Dup15q presents with a wide range of symptoms, including global developmental delay, intellectual disability, neonatal hypotonia, language and motor skill deficits, autism spectrum disorder (ASD), and drug-resistant epilepsy, the latter being associated with an increased risk of sudden unexpected death in epilepsy (SUDEP). Epilepsy is among the most disabling manifestations, affecting over 60% of patients and often presenting with polymorphic seizures that may evolve toward a Lennox–Gastaut–like phenotype.

Electroencephalographic findings may include focal spikes, generalized spikes, diffuse fast rhythms, and hypersarrhythmia. Mild dysmorphic features such as an upturned nose, epicanthal folds, and downslanting palpebral fissures may also be observed [1–5].

Diagnosis begins with a high index of clinical suspicion upon recognition of these signs. Due to phenotypic overlap with other genetic disorders such as Angelman syndrome or Prader–Willi syndrome, high-resolution genetic tools are required to confirm the diagnosis. Chromosomal microarray or comparative genomic hybridization (CMA or array-CGH) is considered the first-line test, as it allows detection of sub-microscopic duplications and provides precise localization and sizing of the abnormality. This recommendation has been endorsed by multiple international guidelines, including those from the American College of Medical Genetics and Genomics [6–10].

Once a duplication is detected, complementary techniques are recommended to determine the underlying mechanism. Fluorescence in situ hybridization (FISH) and conventional cytogenetic studies can detect the presence of *idic(15)* chromosomes and identify potential mosaicism. Additionally, MLPA (Multiplex Ligation-dependent Probe Amplification) is useful for quantifying duplications or triplications in the 15q11.2–q13.3 region. Finally, DNA methylation analysis is essential to determine the parental origin of the duplication, as maternally derived duplications are associated with a more severe clinical phenotype [6,8,11,12].

In terms of treatment, early detection of the duplication allows implementation of personalized interventions aimed at optimizing neurocognitive development and improving management of comorbidities such as refractory epilepsy. Moreover, patient-derived cellular models, such as induced pluripotent stem cells (iPSCs), have opened new avenues for investigating the molecular mechanisms of Dup15q, with the goal of developing targeted therapies tailored to the specific needs of this population [13].

Case Presentation

The patient is a 31-year-old male evaluated by the genetics service. He was born to a 34-year-old mother and a 35-year-old father at the time of conception. The mother had a history of one prior spontaneous abortion. The pregnancy occurred without known exposure to teratogenic agents. Delivery was via cesarean section at 42 weeks of gestation due to post-term pregnancy and was complicated by a nuchal cord, meconium-stained amniotic fluid, and clinical signs of perinatal asphyxia. The patient required orotracheal intubation at birth, although no evidence of meconium aspiration was observed.

Parental consanguinity was not reported. The family history revealed two paternal uncles with unspecified epilepsy and a maternal uncle with an unspecified behavioral disorder. There was no documented family history of congenital

malformations, genetic disorders, behavioral syndromes, or major psychiatric conditions.

Since early childhood, the patient has exhibited global developmental delay, with complete dependence for basic daily activities. Epileptic seizures began at six years of age, predominantly during sleep, and were characterized by generalized tonic features, sialorrhea, sphincter relaxation, and postictal drowsiness. The frequency and severity of seizures progressively increased, with poor response to multiple antiepileptic drugs, resulting in a clinical picture consistent with refractory epilepsy.

Physical examination revealed no overt facial dysmorphisms. The patient could walk independently with a stable gait, without the need for assistive devices. Limited eye contact was noted, as well as forward-facing ears, absence of verbal language with guttural sound production, hand flapping, and motor stereotypies involving the upper limbs and head. No spinal deformities or limb asymmetries were found. External genitalia were normal; the prepuce was redundant with a constriction ring at the glans level, and both testes were located in the scrotum. Macroorchidism was not observed.

Given the clinical constellation of refractory epilepsy, autism spectrum disorder, and intellectual disability, a genetic etiology was strongly suspected. Therefore, molecular analysis was conducted using array-based comparative genomic hybridization (array-CGH) integrated with single nucleotide polymorphism (SNP) analysis. The test was performed using the SurePrint G3 Human CGH+SNP 4×180K platform (Agilent Technologies). This methodology enables the simultaneous detection of genomic copy number variations—such as deletions (losses), duplications (gains), and unbalanced chromosomal rearrangements—thus offering high-resolution insight into the genomic architecture underlying neurodevelopmental disorders.

The analysis identified a heterozygous intersti-

tial duplication of 10.03 megabases (Mb) in the 15q11.2–q13.3 region (genomic coordinates chr15:22572809–32607357, GRCh38 reference genome) (**Figures 1 and 2**). This duplicated segment encompasses several functionally significant genes, including UBE3A, GABRB3, GABRA5, SNRPN, and NDN, all of which play critical roles in neurodevelopment and GABAergic neurotransmission. The 15q11–q13 duplication syndrome (OMIM #608636) has been associated with a spectrum of neurodevelopmental and neuropsychiatric conditions, such

Figure 1. Schematic representation of the pathogenic duplication identified by array-CGH.

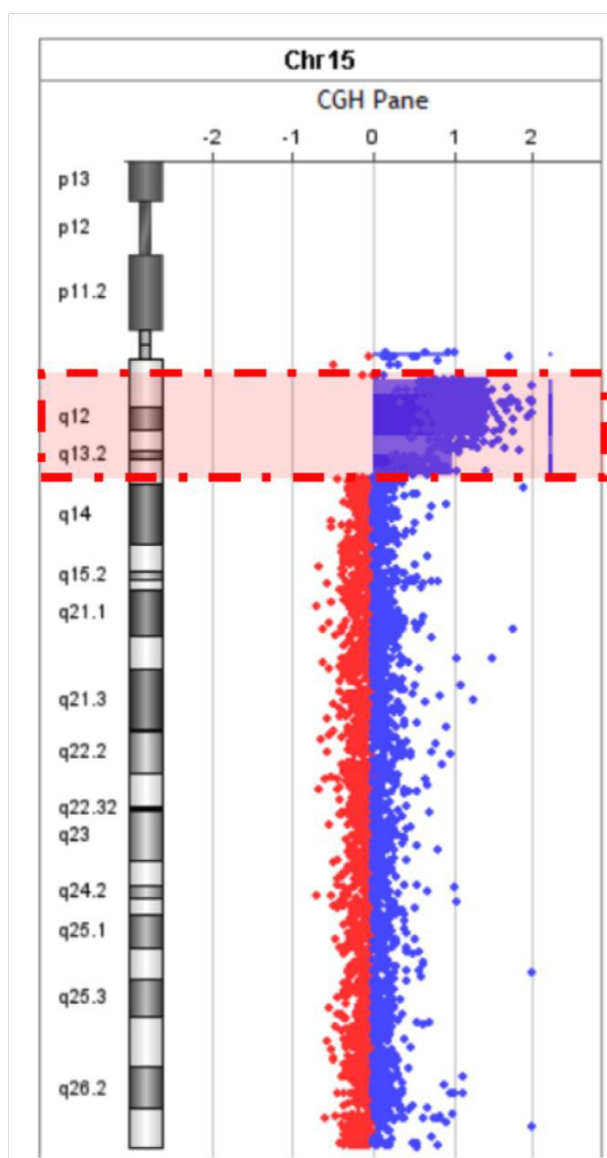
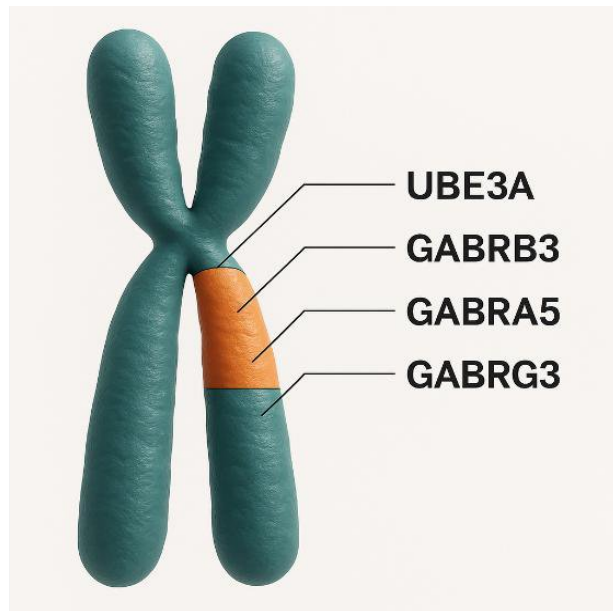


Figure 2. Schematic representation of the pathogenic duplication identified (Duplication 15q11.2-q13.3).



as autism spectrum disorder, intellectual disability, hypotonia, ataxia, seizures, developmental delay, behavioral disturbances, and schizophrenia (PMID: 21324950).

There is growing evidence of a parent-of-origin effect, with maternally inherited duplications more commonly linked to clinical phenotypes. In contrast, paternally derived duplications have been frequently observed in unaffected individuals, although some cases have been documented in patients with autism spectrum disorder (PMID: 23495136) and seizures (PMID: 23633446). However, these findings remain inconsistent, and the current number of reported cases is insufficient to conclusively establish a genotype–phenotype correlation for paternally inherited duplications (PMID: 18840528).

Based on American College of Medical Genetics and Genomics (ACMG) guidelines, this copy number variant is classified as pathogenic. **Table 1** summarizes the characteristics of the genes according to the Human Phenotype Ontology.

Identification of this duplication confirmed the diagnosis of 15q11–q13 duplication syndrome (Dup15q). To further investigate the genetic origin, targeted MLPA analysis of the 15q11–q13 region was performed in both parents, with negative results. This finding supports a de novo origin of the duplication and confirms a non-inherited genetic etiology, based on the observed correlation between genotype, endotype, and phenotype.

This diagnosis enabled precise clinical classification, informed the therapeutic management strategy, allowed for a more accurate prognosis, and laid the foundation for comprehensive genetic counseling for the family.

Discussion

Current literature on 15q11.2–q13.3 duplication syndrome (Dup15q) highlights the importance of early and accurate genetic diagnosis due to its broad phenotypic spectrum and overlap with other neurodevelopmental disorders, such as Angelman syndrome and Prader–Willi syndrome. In the present case, multiple clinical features—including global developmental delay, hypotonia, refractory epilepsy, autism spectrum disorder, and mild dysmorphic features—prompted a comprehensive genetic evaluation that led to confirmation of the diagnosis via array-CGH.

This diagnostic approach is consistent with findings by Bisba et al., who recommend chromosomal microarray (CMA) as a first-line test for detecting copy number variations in patients with complex phenotypes. In their study, over 50% of individuals diagnosed with Dup15q presented with intellectual disability, language impairments, and autistic features, all of which were also observed in this patient [1].

Similarly, Rabeya Akter Mim et al., in a cohort of 260 children with neurodevelopmental disorders, found that 3% had pathogenic duplications in the 15q11–q13 region. These cases exhibited comparable clinical profiles, reinforcing

Table 1. Key Genes Involved in 15q11-q13 Duplication Syndrome: HPO-Based Overview.

Gen	Synonyms	Definition	Disease Associations
UBE3A NCBIGene: 7337 Gene Location: 15q11.2	<i>ANCR,</i> <i>AS,</i> <i>E6-AP,</i> <i>EPVE6AP,</i> <i>HPVE6A,</i> <i>PIX1</i>	This gene encodes an E3 ubiquitin-protein ligase, which is part of the ubiquitin protein degradation system. This imprinted gene is maternally expressed in the brain and biallelically expressed in other tissues. A maternally inherited deletion of this gene causes Angelman Syndrome, which is characterized by severe motor and intellectual retardation, ataxia, hypotonia, epilepsy, absence of speech, and characteristic facies.	15q11q13 microduplication syndrome, Angelman syndrome due to maternal 15q11-q13 deletion, Angelman syndrome due to paternal uniparental disomy of chromosome 15, Angelman syndrome due to an imprinting defect in 15q11-q13, Angelman syndrome due to a point mutation
GABRB3 NCBIGene:2562 Gene Location: 15q12	<i>DEE43,</i> <i>ECA5,</i> <i>EIEE43</i>	This gene encodes a member of the ligand-gated ionic channel family. The encoded protein is one of the subunits of a multi-subunit chloride channel that serves as the receptor for gamma-aminobutyric acid, a major inhibitory neurotransmitter of the mammalian nervous system. This gene is located on the long arm of chromosome 15 in a cluster with two other genes encoding related subunits of this family. This gene may be associated with the pathogenesis of several disorders, including Angelman syndrome, Prader-Willi syndrome, no syndromic orofacial clefts, epilepsy, and autism. Alternatively spliced transcript variants encoding distinct isoforms have been described. [Provided by RefSeq, July 2013]	Epilepsy, childhood absence, childhood absence epilepsy, Lennox-Gastaut syndrome, Epileptic encephalopathy, and early infantile
GABRA5 NCBIGene:2558 Gene Location: 15q12	<i>DEE79,</i> <i>EIEE79</i>	GABA is the major inhibitory neurotransmitter in the mammalian brain where it acts at GABA-A receptors, which are ligand-gated chloride channels. Chloride conductance of these channels can be modulated by agents such as benzodiazepines that bind to the GABA-A receptor. At least 16 distinct subunits of GABA-A receptors have been identified. Transcript variants utilizing three different alternative non-coding first exons have been described. [provided by RefSeq, July 2008]	Non-specific early-onset epileptic encephalopathy, epileptic encephalopathy, early infantile.
SNRPN NCBIGene:6638 Gene Location: 15q11.2	<i>HCERN3,</i> <i>PWCR, RT-LI,</i> <i>SM-D, SMN,</i> <i>SNRNP-N,</i> <i>SNURF-</i> <i>SNRPN, Sm-N</i>	This gene is located within the Prader-Willi Syndrome critical region on chromosome 15 and is imprinted and expressed from the paternal allele. It encodes a component of the small nuclear ribonucleoprotein complex, which functions in pre-mRNA processing and may contribute to tissue-specific alternative splicing. Alternative promoter use and alternative splicing result in a multitude of transcript variants encoding the same protein. Transcript variants that initiate at the CpG island-associated imprinting center may be bicistronic and also encode the SNRPN upstream reading frame protein (SNURF) from an upstream open reading frame. In addition, long spliced transcripts for small nucleolar RNA host gene 14 (SNHG14) may originate from the promoters at this locus and share exons with this gene. Alterations in this region are associated with parental imprint switch failure, which may cause Angelman syndrome or Prader-Willi syndrome. [Provided by RefSeq, Mar 2017]	Prader-Willi syndrome due to an imprinting mutation, autism susceptibility 1, Angelman syndrome, Prader-Willi syndrome due to maternal uniparental disomy of chromosome 15, Angelman syndrome due to imprinting defect in 15q11-q13, Prader-Willi syndrome due to paternal deletion of 15q11q13 type 1, Prader-Willi syndrome due to paternal deletion of 15q11q13 type 2, Prader-Willi syndrome due to translocation.
NDN NCBIGene:4692 Gene Location: 15q11.2	<i>HsT16328,</i> <i>PWCR</i>	This intronless gene is located in the Prader-Willi syndrome deletion region. It is an imprinted gene and is expressed exclusively from the paternal allele. Studies in mouse suggest that the protein encoded by this gene may suppress growth in postmitotic neurons. [Provided by RefSeq, July 2008]	Prader-Willi syndrome due to imprinting mutation, Prader-Willi syndrome due to an maternal uniparental disomy of chromosome 15, Prader-Willi syndrome due to paternal deletion of 15q11q13 type 2, Prader-Willi syndrome due to paternal deletion of 15q11q13 type 1.

ing the diagnostic value of genetic testing not only as a diagnostic tool but also as a guide for clinical management and family counseling [14].

One of the most challenging aspects of this case was refractory epilepsy, a common finding in Dup15q syndrome. Studies such as Elamin et al. (2023) have proposed a molecular basis for this manifestation, demonstrating alterations in sodium channel inactivation in patient-derived cellular models. This evidence supports the development of targeted therapies, which may offer more effective treatment alternatives in the future [2].

Regarding genetic diagnosis, various authors have advocated for systematic integration of chromosomal microarray (CMA) in the initial evaluation of patients with autism, intellectual disability, and epilepsy [6,10]. In this patient, array-CGH precisely established the underlying etiology, confirming its clinical utility as a high-resolution diagnostic tool. Its ability to detect submicroscopic duplications with high sensitivity enabled accurate identification of the structural alteration responsible for the clinical phenotype, consolidating its role as a cornerstone of modern medical genetics.

Furthermore, Bisba et al. emphasized the importance of methylation analysis to characterize duplications in the 15q11–q13 region, particularly to determine parental origin. This factor is prognostically relevant, as maternal duplications are associated with more severe clinical phenotypes. Although this information was not available in the present case, it remains essential for genetic counseling and family planning [1].

In summary, this clinical case illustrates how detailed clinical evaluation, complemented by molecular diagnostic tools such as array-CGH, enables precise identification of an underlying genetic cause and facilitates comprehensive patient management. The experience described aligns with current literature and underscores the need to promote access to molecular cyto-

genetic studies in clinical settings with high suspicion of genomic anomalies.

Conclusion

The 15q11.2–q13.3 duplication syndrome represents a diagnostic challenge in pediatrics due to its clinical heterogeneity and phenotypic overlap with other neurodevelopmental disorders. In the present case, the presence of refractory epilepsy, autism spectrum disorder, and intellectual disability prompted investigation of an underlying genetic cause, which was confirmed through chromosomal microarray analysis (array-CGH).

Application of this technique enabled identification of a pathogenic submicroscopic duplication in the 15q11.2–q13.3 region, thereby establishing the etiologic diagnosis and facilitating a more targeted clinical approach. Array-CGH proved to be a high-resolution method with significant diagnostic value in patients with complex neurological phenotypes, as it allows precise detection of copy number variations that are undetectable by conventional methods.

This case underscores the need to integrate molecular cytogenetic studies into the initial evaluation of patients with high clinical suspicion, as well as the importance of early diagnosis in guiding therapeutic decisions, anticipating prognosis, and providing appropriate genetic counseling to families. Personalized medicine, grounded in molecular understanding of disease, is becoming a fundamental pillar in the management of neurodevelopmental disorders.

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